APEX/PROTEUM3 Converting frames to Bruker format



- Mar (Rayonix)
 - Image plate
 - CCD
- ADSC
- Dectris
 - Pilatus
 - Eiger



APEX/PROTEUM3 Converting frames to Bruker format

- Click on the "Unwarp and convert images" icon from the "Reduce Data" menu
- This will bring up the conversion window
- In older versions of PROTEUM, the plugin was in the instrument menu





APEX/PROTEUM3 Converting frames to Bruker format

Fill in the following parameters and then click the "Convert" button to convert the frames

- Images from: select the image files to convert
- Output format: select the .sfrm format
- Unwarp: this box should be unchecked
- Write to folder: select a output folder
- Header input:
 - Unchecking the missing header box will display all the header parameters
 - Read in the appropriate .hdr file using the icon circled
 - At this point the only things you might have to change are the beam center and detector offset

Images from C:\frames\gu	uest\JK
Output format sfm	(30 image files in 1 run)
Write to folder C:\frames\gu	Jest \JK
Show only missing header	r items
Detector ID	PILATUS-6MF
Detector circular?	No 🔻
 Horizontal tiles 	1
Vertical tiles	1
Pixel width [mm]	0.172
Pixel height [mm]	0.172
Horizontal beam center	3.27
Unit	millimeters 🔻
Origin	center 💌
Direction	right 🔻
Vertical beam center	3.10
Unit	millimeters 💌
Origin	center 🔻
····· Direction	up 🔻
Beam tilt [°]	0.00
Detector pitch angle [°]	0.00
Detector roll angle [°]	-90.00
Detector yaw angle [°]	0.00
Baseline offset	0 🗸
	Convert

C:\frames\guest\training Images from (4 image files in 1 run) Output format sfm unwaro Write to folder C:\frames\guest\training Show only missing header items Detector ID pilatus6m Detector circular? No 🔻 Horizontal tiles 5 Vertical tiles 12 Pixel width [mm] 0.1720 Pixel height [mm] 0.1720 Horizontal beam center 0.09 Unit millimeters 🔻 Oriain center -Direction right 🔻 Vertical beam center -0.09 Unit millimeters -Origin center -Direction up 🔻 Beam tilt [°] 0.00 Detector pitch angle [°] 0.00 90.00 Detector roll angle [°] 0.00 Detector vaw angle [°] Baseline offset 0 Overflow 496476 Electrons/ADU 1.00 * Electrons/photon 1 00 Phosphor efficiency 1.00

Convert

BRUKE

The header file contains all the relevant information for the data frame and can be updated for a particular experimental setup

APEX/PROTEUM3

Header files

- The parameter is fixed when it is updated by clicking on the current value (for example "Detector ID") and inputting the new value
- The checked box indicates a saved parameter
- After all the changes are made, the hardware file can be saved by clicking the circled icon
- This file can be used for every data set collected with this experimental setup
- For beamline data, the detector roll defines the orientation of the spindle and is not always in the header. It will either be 0, 90, 180 or 270. For example, if the spindle is horizontal as with Dectris setups, the roll would be either 90 or 270 (-90). The best way to determine the correct value is to try and index a few degrees of data not just one frame

APEX/PROTEUM3 Header files

- For the header files you need to fill in the information for the detector charcteristics
 - Electrons/ADU
 - Electrons/photon
 - Phosphor efficiency
 - Read noise
- Update the Monochromator for beam polarization
 - Goniometer vertical 2T=90.00 Roll=90
 - Goniometer horizontal 2T=90.00 Roll=0
- Update Start and End positions if necessary
 - Set the geometry
 - Bruker (Eulerian)
 - Bruker (Kappa)
 - Nonius (Eulerian)
 - Nonius (Kappa)
 - Horizontal
 - Mar DTB



Baseline offset	0	
Overflow	496476	- [_]
Electrons/ADU	1.00	-
Electrons/photon	1.00	-
Phoenbor efficiency	1.00	-
Read poise [electrone]	0.00	-
Wavelength [Å]	0.00	-
Manachromator 2Theta [°]	0.07000	-
Monochromator cell [9]	0.00	_
	0.00	-
Exposure time [s]	0.20	
Number of exposures	N	-
Start position	Nonius (Eulenan) 🔻	-
Distance [mm]	400.0000	_
···· 2Theta [°]	0.0000	
···· Omega [°]	0.0000	
Phi [°]	10.0000	
Chi [°]	0.0000	
End position	Nonius (Eulerian) 🔻	
···· Distance [mm]	400.0000	
2Theta [°]	0.0000	
Omega [°]	0.0000	
Phi [°]	11.0000	
Chi [°]	0.0000	- 🛏
Distance offset [mm]	0.0000	
		- Ľ

Pilatus 6M

APEX/PROTEUM3 Making an Active Pixel Mask



For the detectors with sensor gaps, you will want to create a mask to remove both the gaps and beamstop shadow from the image. If the goniometer is a fixed spindle, the beamstop is usually placed along the rotation axis and the shadow may not need to be removed since it's in the Lorentz region

- Load a converted frame into PROTEUM/APEX Viewer
- First adjust the contrast, if needed, so that it's easier to see the gaps. The colored background sets the gaps to black by default which is the color needed for generating a mask



APEX/PROTEUM3 Making an Active Pixel Mask



- Use a graphics program convert the active areas to white and the inactive areas to black and save a PNG file adjusting the image size to match actual frame size In a cmd window type:
 - Bnrun c:\bn\src\misc\imagesum.pyc *filename*.png to mask._am
 - You must run imagesum from the same folder as the PNG file
 - This will convert the PNG file to a mask file in sfrm format
 - The *filename_xa_01_0001*.sfrm convention can also be used as output







Initial PNG file

Image with updated colors

Final mask

APEX/PROTEUM3 Making an Active Pixel Mask



- Use a graphics program convert the active areas to white and the inactive areas to black and save a PNG file adjusting the image size to match actual frame size
- Typical Pilatus 6M image is 2463 x 2527 but the png file outputted from PROTEUM will be much smaller so you need to change the image resolution of the png file to match that of the actual image otherwise you will get an error in SAINT

training.png - IrfanView	
File Edit Image Options View Help	
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	el co

Resize/Resample image	X		
Current size: 762 x 782 Pixels New size: 2463 x 2527 Pixels	Some standard dimensions (pixels): New size: © 640 x 480 © 1280 x 720 (HD)		
Set new size: Width: 2463	800 x 600 1920 x 1080 (FHD) 1024 x 768 3840 x 2160 (4k) Best fit to desktop 7680 x 4320 (8k) Desktop size (no aspect ratio) Haff Swap sides Add to standard box		
Preserve aspect ratio (proportional) Apply sharpen after Resample Adjust DPI based on new sizes (asp. ratio) DPI: 96 (auto calc. for cm/inches)	Size method: Resample (better quality), use Filter: Lanczos (slowest) Resize (faster, lower quality) Use fast Resample filter for image shrinking Try to improve gamma for Resample		
ОК	Cancel		



APEX/PROTEUM3 Reading an Active Pixel Mask

If the standard mask filename convertion is not used, you will have to read in the Pre-existing mask

- Under Integration options
 - Select more options
 - Click on "Use Preexisting static mask"
 - Browse for the mask file

Sample Instrument	Chart 1	Windows Help			Integrate Images _ 🗗 🗙 BROKE
🗎 📦 🍯 📑	8	\$2			
Set Up	/ Setup	1			
Evaluate		Starting Image Filename	Images	Output Filename	
Collect	1	C:\frames\guest\JK\cy31_01_0001.sfm	30	C:\frames\guest\jk\work\cy31_01.raw	Resolution Limit [Å]: 2.200
Reduce Data	2				
	3				Unit Cells:
R	4				a=107.06Å, α= 90.00°, V=2546839Å ³ b=107.06Å, β= 90.00°, Bhombohadral R
\hat{Y}^{α}_{β}	5				c=256.60Å, y=120.00°
JIZY P	6	Integration Options		? ×	
Integrate Images	7				
	8	Model Profiles	Background	Jpdate	
	9	✓ Enable LS Profile Fitting	Ba	ckground Update Scaling Factor: 1.000	
ene ene	10	Blend Profiles from All Detector Regions			
Index Crystal Faces	11	Intensity/Sigma Lower Limit for Model Profile Update: 10.000	inage ordeue		
	12	Fraction of Model Profile Maximum for Simple Sum Mask: 0.050	Active In	nage Queue Half-Width [Images]: 7	
	13	Intensity/Sigma Upper Limit for LS Model Profile F#	-Beam Monito		
	14	Laura Darak dia Lini ka LC Madal Parila Di Italia na			
	15	Lower Resolution Limit for L3 Model Prolife Rit (A). 9995.000		Enable Beam Monitor Normalization	
Scale	16	Profile XYZ Half-Widths: 4 4 4		Normalize each Run Separately	
	17	Active Mask	Twin Overlap	Determination	
	18			Minimum Common Victoria (%)	-
	19				-
	20	Fractional Lower Limit of Average Intensity:		Separation Factor: 1.000	
Unwarp and Convert Images	21	Use Pre-Existing Static Mask:		Maximum Range: 1.300	-
	22	Active Mask File: C:\frames\guest\JK\maskam	-Modulated St	ucture Integration	-
	24	Ilea Pre Scietine Deservie Meder		Maximum Satellite Index:	-
	25				
	26	Algorithm	Output / Diag	nostic Files	
	27	Use Narrow Frame Algorithm Use Wide Frame Algorithm		Generate Diagnostic Plot Files	
	28			Keep Temporary Files	
	29	Monte Carlo Simulation		Annend Listing Files	Refinement Options
	30	Number of Monte Carlo Simulations:		Hide Log Window	
	31				Integration Options
	32	Image Timeout		Verbosity of Listing File: 2	Cad Data
	33	Wait for Images During Data Collection	C	abat Ordnut Engranger (Impaga): 100	Hind Huns
	34	E Tractor integer calling bata collectori	Snap	snor outpur nequency [images].	Import Runs from Experiment
Evamina Data	35	Fewer Options		OK Cancel	
Find Structure	36				
Report	37				Start Integration



APEX/PROTEUM3 Updating the Beam Center

Sometimes the beam center is not correctly stamped in the frame header. To change this:

- Activate the rectangle tool and zoom in the image to view the beamstop shadow.
- Using the circle tool, drag the circle down until it's very small with the left mouse button. This will show the position of the beam center based on the values in the frame header.
- Click the manual button for the corrections. The boxes for the X and Y offset will become active.



APEX/PROTEUM3 Updating the Beam Center



- Enter new values for the offsets, as you do the circle will move.
- When the circle is where you think the beam is, continue with indexing. The new values for the offsets will be used.
- You can update the beam center in the frame headers by inputting the new values in "Unwarp and Convert" and converting the frames again.



APEX/PROTEUM3 Photon Countering Detectors



- If the diffraction is not particularly strong, there are a couple of options to minimize harvesting bogus reflections from the active area/gap boundary
 - Add the line "scalebeforeharvest=0" to the c:\bn-config.py (windows), /usr/local/lib/bn-config.py (Linux) file. It can go anyway in this file. This will slow things down a bit during harvesting
 - Check the "smooth images" box in the "Harvest spots" window
 - Check the "Save reflections that only span images" box in the "Harvest spots" window
 - Remove the bad reflections with RLATT

APEX/PROTEUM3 Writing out CBF frames during experiment



You can automatically write out CBF frames along with sfrm frames during data collection by setting the AutoCBF flag in the BIS configuration file.

- Shutdown BIS
- Edit the BrukerAXS.Server.BIS.exe.config found in the c:\Program files (x86)\Bruker AXS\DIFFRAC.MEASUREMENT CENTER.V6 folder
- Search for CBF in the file, set the CBF flag to "True" and save the file:
 - <!-- Automatically convert Bruker frame files to CBF frame files -->

```
<!-- Default False: Only create Bruker Frame Files -->
```

```
<setting name="CBF" serializeAs="String">
```

```
<value>False</value>
```

```
</setting>
```

<!-- Automatically convert Bruker frame files to CBF frame files -->

```
<!-- Default False: Only create Bruker Frame Files -->
```

```
<setting name="CBF" serializeAs="String">
```

```
<value>True</value>
```

```
</setting>
```

Restart BIS



APEX/PROTEUM3 XDS, Convert sfrm images to cbf format

To use XDS to process Bruker images you first have to convert our sfrm format to CBF.

- Convert .sfrm images to CBF format using "Unwarp and Convert images" under the Reduce Data menu
- Assign the scans to convert in the "Images from" box using the browser button
- Select the "Output format" as CBF
- Leave the unwarp box unchecked
- Assign an output folder in the "Write to folder" box
- Click the Convert button



APEX/PROTEUM3 Using XDS for data reduction



- Run the genererate_xds.inp script on each data run to create the XDS input files:
 - generate_xds.inp "filename_01_???.cbf"
- Run XDS and XSCALE as normal
- Note that the Y beam position is inverted
 - X=514, Y=510, Y will be 514 in the script



Innovation with Integrity

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