Selective base excision repair of DNA damage by the non-base-flipping DNA glycosylase AlkC

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Abstract

DNA glycosylases preserve genome integrity and define the specificity of the base excision repair pathway for discreet, detrimental modifications, and thus, the mechanisms by which glycosylases locate DNA damage are of particular interest. Bacterial AlkC and AlkD are specific for cationic alkylated nucleobases and have a distinctive HEAT-like repeat (HLR) fold. AlkD uses a unique non-base-flipping mechanism that enables excision of bulky lesions more commonly associated with nucleotide excision repair. In contrast, AlkC has a much narrower specificity for small lesions, principally N3-methyladenine (3mA). Here, we describe how AlkC selects for and excises 3mA using a non-base-flipping strategy distinct from that of AlkD. A crystal structure resembling a catalytic intermediate complex shows how AlkC uses unique HLR and immunoglobulin-like domains to induce a sharp kink in the DNA, exposing the damaged nucleobase to active site residues that project into the DNA. This active site can accommodate and excise N3-methylcytosine (3mC) and N1-methyladenine (1mA), which are also repaired by AlkB-catalyzed oxidative demethylation, providing a potential alternative mechanism for repair of these lesions in bacteria.

Keywords 3-methyladenine; 3-methylcytosine; base excision repair; DNA glycosylase; DNA repair

Subject Categories DNA Replication, Repair & Recombination; Structural Biology

DOI 10.15252/embj.201797833 | Received 22 July 2017 | Revised 11 September 2017 | Accepted 22 September 2017 | Published online 20 October 2017


Introduction

Cellular metabolites and environmental toxins damage DNA by creating a variety of covalent DNA adducts that impair normal cellular processes and lead to heritable diseases, cancer, aging, and cell death (Friedberg et al., 2006; Jackson & Bartek, 2009). Alkylation of nucleobase substituents and ring nitrogens is a major source of DNA damage that can directly inhibit replication and transcription or can degenerate to other forms of damage including abasic sites and strand breaks (Wyatt & Pittman, 2006; Krokan & Bjørkás, 2013). The toxicity of these DNA lesions is the basis for the use of alkylating agents in cancer chemotherapy (Sedgwick, 2004). To avoid the toxic effects of DNA alkylation and maintain integrity of their genomes, all organisms possess multiple repair mechanisms to remove the diversity of alkyl-DNA modifications. Nucleotide excision repair (NER) is the predominant mechanism to eliminate helix-distorting and bulky adducts, whereas small nucleobase modifications (e.g., methyl, etheno groups) are repaired by direct reversal or base excision repair (BER) pathways (Sedgwick, 2004; Reardon & Sancar, 2005; Mishina & He, 2006). N1-methyladenine (1mA) and N3-methylcytosine (3mC), predominant in single-stranded (ss) DNA and RNA, are demethylated by the AlkB family of Fe(II)/α-ketoglutarate-dependent dioxygenases. In contrast, N3- and N7-alkylpurines, the most abundant double-stranded DNA alkylated products, are removed from the genome by BER, whereby the damaged nucleobase is cleaved from the phosphodeoxyribose backbone by lesion-specific DNA glycosylases that hydrolyze the N-glycosidic bond. The resulting abasic site is nicked by apurinic/apyrimidinic (AP) endonuclease to generate a 3'-hydroxyl substrate for gap repair synthesis by DNA polymerase.

DNA glycosylases that remove alkylation damage are essential to eukaryotes, archaea, and bacteria (Riazuddin & Lindahl, 1978; Thomas et al., 1982; Chen et al., 1989; O’Connor & Laval, 1991; Birikeland et al., 2002). Bacteria often contain paralogs that eliminate diverse types of damage resulting from endogenous versus environmental sources. For example, in Escherichia coli, the AlkA glycosylase is induced to remove a broad spectrum of alkylated and deaminated bases upon cellular exposure to alkylation agents, while the constitutively active Tag enzyme is highly specific for N3-methyladenine (3mA) formed endogenously (McCarthy et al., 1984; Bjelland et al., 1993, 1994; Saparbaev et al., 1995; O’Brien & Ellenberger, 2004). Damage recognition and excision by these and other glycosylases rely on a base-flipping mechanism, whereby the damaged nucleotide is rotated ~180° around the phosphate backbone and sequestered inside a nucleobase binding pocket on the protein surface (Brooks et al., 2013). This pocket contains residues that facilitate depurination by activating or stabilizing the nucleobase leaving group and/or electrostatically stabilizing an oxocarbenium intermediate that reacts with a water molecule to generate the abasic site...
product (Fig 1A) (Drohat & Maiti, 2014). The extrahelical orientation of the substrate in the catalytic complex is stabilized by DNA-intercalating protein residues that fill the space generated by the missing base. Thus, the specificity of a DNA glycosylase for a particular substrate is achieved by a combination of duplex interrogation and complementarity of the nucleobase and the active site pocket.

The bacterial AlkC and AlkD glycosylases, originally identified in Bacillus cereus (Bc), comprise a distinct superfamily of DNA glycosylase selective for positively charged N3- and N7-alkylpurines (Alseth et al., 2006; Dalhus et al., 2007). They specifically lack excision activity toward uncharged 1,N6-ethenoadenine or hypoxanthine nucleobases (Alseth et al., 2006), the common substrates of many previously characterized alkylpurine DNA glycosylases. Structures of BcAlkD revealed a new fold composed of tandem HEAT-like repeats that form a left-handed solenoid around the DNA duplex (Rubinson et al., 2008, 2010; Rubinson & Eichman, 2012). Unlike other glycosylases, AlkD does not use a base-flipping mechanism for damage recognition or base excision (Mullins et al., 2015b). Instead, the protein traps non-Watson–Crick base pairs in a sheared, base-stacked conformation with catalytic active site residues in direct contact with the deoxyribose, but not the nucleobase, of the damaged nucleoside. The lack of protein–nucleobase contacts enables AlkD to excise both the major and minor groove lesions 3mA and N7-methylguanine (7mG). Similarly, by not confining the modified nucleobase inside a binding pocket, AlkD can excise bulky lesions, including pyridyloxobutyl (POB) adducts resulting from the carcinogen NNK (nicotine-derived nitrosamine ketone) (Rubinson et al., 2010), as well as N3-yatakemycinyladenine (YTMA) formed from the highly genotoxic bacterial natural product yatakemycin (YTM) (Xu et al., 2012; Mullins et al., 2015b, 2017). Thus, AlkD excises a diverse spectrum of cationic lesions, including bulky lesions expected to be processed by NER.

AlkC from B. cereus (BcAlkC) shares 15.8% identity and 33.7% similarity with BcAlkD and has been predicted to adopt the HLR architecture (Dalhus et al., 2007). However, in contrast to AlkD, AlkC has a strong preference for 3mA and N3-methylguanine (3mG) and only weak activity for 7mG (Alseth et al., 2006; Mullins et al., 2013). Given AlkD’s unique non-base-flipping mechanism and activity for bulky lesions, we were interested to expand our understanding of this new DNA repair superfamily by determining the basis for AlkC’s apparent limited substrate specificity. We carried out a comprehensive phylogenetic, biochemical, and structural comparison of AlkC and AlkD proteins and found that the majority of AlkC proteins contain an immunoglobulin (Ig)-like domain not yet observed in a DNA repair enzyme and that is essential for base excision activity in those enzymes. We also found that unlike AlkD, AlkC does not remove YTMA adducts either in vitro or in vivo. The crystal structure of Pseudomonas fluorescens (Pf) AlkC bound to damaged DNA revealed that the HLR and Ig-like domains wrap almost completely around the DNA duplex to recognize the damaged base pair from opposite major and minor grooves. Like AlkD, AlkC does not extrude the damaged nucleobase from the DNA helix. However, unlike AlkD, AlkC induces a sharp kink in the DNA and accesses the damage by inserting active site residues into the exposed base stack. The structure and supporting mutational analysis of base excision by AlkC provide a mechanistic basis for how AlkC selectively excises 3mA from DNA. Additionally, we show that AlkC’s unique active site is also capable of robust excision activity for 3mC, which is normally repaired by AlkB-catalyzed oxidative demethylation. This work provides a molecular basis for how a non-base-flipping glycosylase selects for discrete methylated bases, and describes an alternative mechanism for removal of 3mC in bacteria.

Results and Discussion

Proteins within two AlkC subgroups are functionally distinct from AlkD

Despite a predicted similarity in HLR secondary structure, AlkC and AlkD from B. cereus were previously shown to be phylogenetically and functionally distinct (Alseth et al., 2006; Mullins et al., 2013). To better understand the AlkC family of proteins and how they differ from AlkD, we obtained 779 AlkC and 764 AlkD orthologous sequences by performing an iterative PSI-BLAST search against the NCBI non-redundant protein database and then used them to
reconstruct a phylogenetic tree. 93% of AlkC sequences were widespread among actinobacteria, bacteroidetes, firmicutes, and proteobacteria, and only rarely found in archaea and eukaryotes, largely consistent with a previous analysis (Backe et al., 2013). We found that AlkC proteins clustered into two distinct clades that we denote AlkCα and AlkCβ (Fig 1B), both of which contain an HLR domain distinct from AlkD mainly within the N-terminal 60–80 residues. The AlkCβ clade, which constitutes the majority (70%) of the total AlkC sequences, contains an additional ~100-residue putative immunoglobulin (Ig)-like fold at the C-terminus (Fig 1C, Appendix Fig S1).

The specificity of AlkC for 3mA has been characterized only from the B. cereus enzyme, which belongs to the AlkCα class (Alseth et al., 2006; Mullins et al., 2013). To determine whether this 3mA selectivity is a general characteristic of both AlkCα and AlkCβ proteins, we cloned and purified five AlkCα and two AlkCβ orthologs and measured their ability to excise 3mA and 7mG nucleobases from N-methyl-N-nitrosourea (MNU)-treated genomic DNA, the major methylated products of which are 7mG (66%) and 3mA (8%) (Lawley, 1976). All seven AlkC proteins released the maximum amount of 3mA after 1 h as judged by treatment of substrate with HCl, but exhibited only very low activity toward 7mG as compared to an AlkD control (Fig 1D). Consistent with this, both BcAlkC (α) and PfAlkC (β) excised 7mG from a defined oligodeoxynucleotide substrate after 24 h, albeit with much slower kinetics than BcAlkD (Appendix Fig S2A). Thus, 3mA specificity seems to be a general property of all AlkC proteins, independent of the presence of the putative Ig-like domain.

We also found that AlkC’s specificity for N3-alkyladenine adducts is limited to small adducts. Neither BcAlkC nor PfAlkC was able to cleave YTMAs lesions for which AlkD exhibits robust activity (Mullins et al., 2017) (Appendix Fig S2B). We previously showed that in addition to AlkD excision of YTMAs in vitro, Bacillus anthracis cells lacking AlkD exhibited a sensitivity to YTM (Mullins et al., 2015b). We therefore tested growth of AlkC-deficient B. anthracis strains in the presence of YTM. Consistent with our in vitro results, YTM sensitivity of a ΔalkC strain did not differ from that of wild-type B. anthracis, nor did a ΔalkC/alkD double mutant show additional sensitivity compared to ΔalkD (Appendix Fig S2C). Thus, substrate specificities of both α- and β-subgroups of AlkC are distinct from AlkD, suggesting that AlkC utilizes a different strategy to recognize damaged DNA despite its predicted structural similarity to AlkD.

AlkC encircles and bends damaged DNA

To determine the structural basis for AlkC’s preference for 3mA and whether a non-base-flipping mechanism is a common feature among HLR glycosylases, we determined a crystal structure of PfAlkC in complex with DNA containing 1’-aza-2’-A’-dideoxyribose (1aR), which mimics the oxocarbenium ion intermediate formed upon nucleobase dissociation (Fig 1A) (Hollis et al., 2000; Chu et al., 2011; Schramm, 2011). The PfAlkC/1aR-DNA model was refined against X-ray diffraction data extending to 1.8 Å resolution to a crystallographic residual of 14.1% (Rfree = 16.4%) (Appendix Table S1) and contained two crystallographically unique protein–DNA complexes, each with 361 of 369 amino acids and all 22 nucleotides clearly defined by the electron density. The structures of the two complexes in the asymmetric unit (asu) were virtually identical (r.m.s.d. = 0.42 Å for all atoms), differing only in the location of a statically disordered 5’-overhanging adenosine A1 on the undamaged strand that forms a crystal packing contact with the same nucleotide in an adjacent protein/DNA complex.

The PfAlkC crystal structure confirmed the presence of distinct HLR and Ig-like domains, which together wrap almost completely around a highly bent DNA duplex (Fig 2). As a consequence of its interactions with the HLR and Ig-like domains, the DNA is sharply bent at the 1aR lesion by 60° away from the minor groove and the minor groove widened by 7 Å. The HLR domain (helices αA’-αN) is similar to that of AlkD in that it contains an N-terminal α-helical bundle (αA’-αC) followed by five HLRs—αD-αE, αF-αG, αH-αI, αL-αK, and αL-αM—that together form a left-handed solenoid that wraps around one-half of the DNA duplex from the minor groove side (Fig 2A and B). The N-terminal α-helical bundle contacts the backbone of the undamaged strand from the minor groove using polar side chains and the helix dipole of helices αA’ and αC, which point directly at thymines T6 (opposite the 1aR) and T7, respectively (Fig 2A). All HLRs except HLR1 directly contact the DNA, primarily along the phosphate backbones. The C-terminal helices of each HLR adorn basic and polar side chains that bind either strand from the minor (HLR2, HLR3) or major (HLR4, HLR5) groove sides (Fig 2C). HLR2 and HLR3 also contain conserved glutamate side chains that directly contact the 1aR. Between HLR3 and HLR4, an 11-residue loop/α-helix insertion (αL-loop) not found in AlkD penetrates the minor groove at the lesion. The HLR domain is tethered by a 9-residue linker to the Ig-like fold, which is composed of nine β-strands (βA1-βG) and an extended loop that contacts the DNA from the major groove side (Fig 2A). Both domains form a positively charged, highly conserved concave surface that engulfs the DNA (Fig 2B). Extensive polar and van der Waals contacts are formed with the DNA backbone 5’ to the 1aR and along the entire undamaged strand (Fig 2C).

The AlkC Ig-like domain is a unique DNA binding motif in bacteria

Ig-like domains are prevalent in both bacteria and eukaryotes as a generic scaffold (Bork et al., 1994; Halaby & Morron, 1998; Halaby et al., 1999; Bodelón et al., 2013) and are important for sequence-specific DNA binding in some eukaryotic transcription factors (Cho et al., 1994; Cramer et al., 1997; Becker et al., 1998; Chen et al., 1998; Nagata et al., 1999; Bravo et al., 2001; Rudolph & Gergen, 2001; Tahirov et al., 2001; Lamoureux et al., 2002; Rohs et al., 2010). To our knowledge, no bacterial Ig-like domains have been reported to bind DNA or to function in DNA repair. The Ig-like domain of PfAlkC is composed of a nine-strand (βA1-βG) antiparallel β-sandwich, in which βA1, βA2, βA3, βB, and βE form one β-sheet packed against a second sheet formed by strands βC, βF, and βG (Fig 2A). The topology is consistent with the C2 subtype (Fig 3B) but with very low sequence similarity, a highly kinked βA strand (βA1-βA2-βA3), and a longer βC strand than other C2-type Ig-like domains (Halaby et al., 1999; Bodelón et al., 2013). The closest structural homolog of AlkC’s Ig-like domain as judged by a Dali search (Holm & Laakso, 2016) is found in a bacterial β-glucosidase and has been proposed to contribute to substrate binding and dimerization of that enzyme (McAndrew et al., 2013).
The Ig-like domains of eukaryotic transcription factors mediate DNA binding through the loops between strands βA-βB and βE-βF (AB- and EF-loops) and the C-terminal tail regions, all of which emanate from the β-sandwich core (Fig EV1) (Cho et al., 1994; Cramer et al., 1997; Becker et al., 1998; Chen et al., 1998; Nagata et al., 1999; Bravo et al., 2001; Rudolph & Gergen, 2001; Tahirov et al., 2001; Lamoureux et al., 2002; Rohs et al., 2010). Although we have no evidence for sequence-specific binding by AlkC, AlkC’s Ig-like domain also mediates DNA binding by the EF-loop, which in PfAlkC contains a conserved Met-Thr-Thr-Arg motif (residues 336–338) that contacts nucleobases on both strands in the major groove (Fig 3A). Side- and main-chain groups within this motif interact with the phosphoribose groups on the undamaged strand immediately 5’ to the orphaned thymidine T6, and Thr337 is engaged with cytosine C5 on the damaged strand. In contrast to the eukaryotic transcription factors, AlkC’s AB-loop does not contact DNA, despite its orientation toward the DNA (Fig EV1). Instead, PfAlkC makes unique DNA binding interactions using the βC strand and the preceding CC’-turn (Fig 3A and B). Side chains of Lys309, Phe311, and Lys312 on the βC’ strand interact with the backbones of both strands, and Lys301 and Ser302 on the CC’-turn form polar interactions with phosphates on the 5’-side of the lesion strand. As a consequence of these contacts to both DNA strands, the Ig-like domain in AlkC plays an important role in stabilizing the kinked DNA conformation toward the major groove.

Consistent with a functional importance of the Ig-like domain, deleting it from PfAlkC (PfAlkCΔC, Fig EV2A and B) abrogated base excision activity (Fig 3C), likely owing to a severe decrease in DNA binding affinity relative to the wild-type protein (Fig EV2C). The dependence of PfAlkC activity on the Ig-like domain is interesting given the absence of this domain in the fully functional AlkCa proteins, which suggests that the AlkCa HLR domain contains a structural feature that compensates for DNA binding in the absence of the Ig-like domain. Perhaps the most distinguishing feature of the AlkCa HLR primary structure is an 8–10 residue insertion within helix αE of HLR1 (Appendix Fig S1). Whereas HLR1 does not contact the DNA in the PfAlkC structure, a homology model of BcAlkC shows that the extra residues likely endow AlkCa proteins...
with a novel DNA binding interaction to the damaged strand (Fig EV3). Together with potential additional interactions between the N-terminal helical bundle and the undamaged strand, these extra HLR1-DNA contacts would help stabilize the DNA kink. Although the precise manner in which AlkC promotes compensation for the lack of the Ig-like domain has yet to be determined, it is clear that DNA binding by the Ig motif is essential for AlkC's function.

AlkC inserts its active site into the DNA duplex in lieu of base flipping

Contacts between the HLR domain and the lesion help to explain AlkC's specificity for 3mA and its mechanism of base excision. At the DNA bend, both the 1aR lesion and its opposing thymidine on the AlkC's specificity for 3mA and its mechanism of base excision. At the Contacts between the HLR domain and the lesion help to explain base flipping. AlkC inserts its active site into the DNA duplex in lieu of the undamaged strand, these extra HLR1-DNA contacts would help stabilize the DNA kink. Although the precise manner in which AlkC promotes compensation for the lack of the Ig-like domain has yet to be determined, it is clear that DNA binding by the Ig motif is essential for AlkC's function.

To gain a better idea of how the protein selects for 3mA, we modeled the nucleotide in the active site using 1aR as a guide (Fig 4C). Holding the deoxyribose ring in a fixed position, the 3mA nucleobase could adopt only a limited range of positions that varied within a 15º torsional rotation about the N-glycosidic bond, constrained by π-stacking against either its 3'-neighbor guanine G8D on one side or the protein surface on the other. Against the protein, the N1-C2 edge of the 3mA nucleobase abuts the indole side chain of Trp164 to form a stabilizing cation-π interaction. In this conformation, the N3-methyl group resides in a pocket formed by Phe122 and Glu121 (Fig 4B). These contacts to the damaged nucleobase explain AlkC's inability to excise bulky minor groove YTMA lesions. Moreover, in this model, the Hoogsteen face of the purine ring is snugly nestled between the flanking nucleobases at the DNA kink, suggesting that purine N7-adducts would be sterically disfavored from this collapsed major groove, providing a rationale for AlkC's low activity toward 7mG.

The position of the 1aR deoxyribose in the crystal structure provides a model for catalysis of base excision by AlkC. The N1 nitrogen of 1aR, which mimics the anomer of C1' carbon in the oxocarbenium intermediate, is positioned perfectly for nucleophilic attack by the water molecule held in place by Glu121, Arg152, and Glu156 (Fig 4A and B). These putative catalytic protein–DNA contacts are made without flipping the modified nucleotide into an active site pocket. In fact, the nucleotide is sterically constrained from further rotation around the DNA backbone. A network of alternating charges between Glu121-Arg152-Glu156-Arg159 spans the backbone in a 3’–5’ direction across the damaged nucleotide. Arg152 and Arg159 electrostatically stabilize the two phosphates flanking the 1aR and hold the carboxylate side chains of Glu121 and Glu156 in close proximity to the 1aR sugar ring (Fig EV4B). Thus, Glu121 and Glu156 are positioned to electrostatically stabilize the positive charge that develops on the deoxynucleobase during base excision and to orient and deprotonate the catalytic water nucleophile for attack of the anomer of C1’ carbon, without the need to flip the damaged nucleobase out of the DNA.
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To validate this region of the protein as the active site, we tested the contribution of Glu121, Glu156, and Trp164 to catalysis by substituting individual residues with alanine and comparing the ability of the mutants to excise 3mA from methylated genomic DNA. We verified that the mutations did not compromise protein integrity or DNA binding (Fig EV2). The Glu121Ala substitution abrogated 3mA excision activity and Glu156Ala significantly decreased 3mA excision relative to wild-type (Fig 4D), consistent with a catalytic role for these residues. In contrast, removal of the indole side chain from Trp164 had no effect on the total amount of 3mA excised after 5 min, suggesting that this residue either does not participate in catalysis or has a less detectable role in this assay. These structural and biochemical data clearly designate the region surrounding the 1aR as the active site and demonstrate that like AlkD, the AlkC protein uses a non-base-flipping mechanism to access its substrate.

DNA glycosylase product complexes are often remarkably similar in conformation to their substrate or intermediate structures, and consequently, these enzymes are typically inhibited by their abasic site products, which we recently showed to be true for the non-base-flipping glycosylase AlkD (Brooks et al, 2013; Mullins et al, 2015b, 2017). In an effort to trap an AlkC product complex, we crystallized PfAlkC with DNA containing a tetrahydrofuran (THF) abasic site mimetic. The PfAlkC/THF-DNA crystal structure was refined to a crystallographic residual of 16.8% (Rfree = 22.5%) against X-ray data extending to 2.4 Å (Appendix Table S1, Fig EV5). The asu of the THF structure contained two protein–DNA complexes with HLR and Ig-like domains wrapping around highly bent DNA duplexes as in the 1aR structure. However, unlike the virtually identical protein–DNA complexes in the 1aR structure, each protein–DNA complex in the THF structure differed significantly in the relative position of their Ig-like and HLR domains as well as in the configuration of the DNA, with a 60° bend angle in one and an 85° bend in the other (Fig EV5G). The two DNA duplexes in the asu were stacked on one another at the overhanging A1 nucleotides on one end, but did not make lattice contacts on the other, and thus, the different conformations likely indicate the flexible nature of DNA near the lesion. Most notably, there was a substantial difference in the positions of THF and 1aR bound to AlkC. The THF moieties were slipped out of the DNA helix as single nucleotide bulges, each adopting a distinct conformation and a set of contacts to AlkC outside of the active site DNA complex in the 1aR structure. However, unlike the virtually identical protein–DNA complexes in the 1aR structure, each protein–DNA complex in the THF structure differed significantly in the relative position of their Ig-like and HLR domains as well as in the configuration of the DNA, with a 60° bend angle in one and an 85° bend in the other (Fig EV5G). The two DNA duplexes in the asu were stacked on one another at the overhanging A1 nucleotides on one end, but did not make lattice contacts on the other, and thus, the different conformations likely indicate the flexible nature of DNA near the lesion. Most notably, there was a substantial difference in the positions of THF and 1aR bound to AlkC. The THF moieties were slipped out of the DNA helix as single nucleotide bulges, each adopting a distinct conformation and a set of contacts to AlkC outside of the active site (Fig EV5D–F). As a consequence, adenine A6\(^{13}\), immediately 5’ to the THF on the damaged (D) strand, formed an opportunistic base pair with thymine T6\(^{13}\) opposite THF on the undamaged (U) strand, leaving T7\(^{13}\) (instead of T6\(^{13}\)) unpaired. The mismatched A6\(^{13}\)-T6\(^{13}\) base pair was observed in both Watson–Crick and Hoogsteen orientations in the two complexes in the asu (Fig EV5F). Because neither THF contacted the active site residues, similar to that observed in a

Figure 4. AlkC inserts its active site into the DNA.
A Close-up view of the AlkC active site (blue) bound to 1aR-DNA (gold). The 1aR and opposite thymine are magenta and green, respectively. Water is shown as a red sphere, and hydrogen bonds are depicted as dashed lines. Composite omit electron density contoured to 1σ is superimposed against only the DNA for clarity. Superscripts in nucleotide labels refer to the 1aR-containing, damaged (D) strand or the opposite, undamaged (U) strand. The pentaerythritol propoxylate molecule that occupies the active site has been omitted for clarity (see Fig EV4B).
B Schematic of the alignment of a catalytic water molecule (blue) against the 1aR oxocarbenium mimetic by AlkC active site residues. Hydrogen bonds are shown in red.
C A hypothetical model for AlkC bound to a 3mA-DNA substrate was generated by superimposing the 3mA deoxyribose ring onto that of 1aR in the crystal structure, followed by rotating about the 3mA χ (N-glycosidic bond) torsion angle to maximize van der Waals interactions. The solvent accessible surface of AlkC is shown as a transparent white envelope.
D Release of 3mA from methylated genomic DNA after a 5-min incubation with either HCl, no enzyme (mock), PfAlkC (WT), E121A, E156A, or W164A. Values are mean ± SD (n = 3).
and product complexes by remaining stacked in the DNA duplex. Previously, we showed that the excised nucleobase stabilizes the conformation of the abasic DNA on its own. Rather, we previously showed that the excised nucleobase stabilizes intermediate complexes containing free 3mA nucleobase were unsuccessful, likely owing to the lack of base stacking at the sharp kink in the DNA, and suggesting that the nucleobase readily dissociates following N-glycosidic bond cleavage.

Structural differences between AlkC and AlkD

With crystal structures for both AlkC and AlkD in hand, we are able to understand the basis for their differences in locating and discriminating DNA alkylation damage. Despite the low sequence conservation between the HLR domains of BcAlkD and PfAlkC (16.3% identity and 27.1% similarity), they adopt the same general C-shaped architecture that complements the DNA helix (Fig 5A–C). Neither protein flips the target nucleobase out of the helix, but they use different mechanisms to interrogate the minor groove for lesion recognition. The modified nucleobase in the BcAlkD complex does not contact the protein, but remains stacked within a DNA duplex that is only modestly bent (Appendix Fig S3), which allows the enzyme to tolerate different positions and sizes of alkyl substituents. The bulky YTMA lesion in particular resides in a cleft between the surface of the protein and the kinked DNA (Fig 5B). This loop occupies the space in the minor groove that would be occupied by YTMA and explains why AlkC is unable to excise bulky minor groove lesions. Because of its direct contact to the modified nucleobase, the presence of the YTMA loop also restricts the size of the lesion that can be accommodated (Fig 4C).

The highly kinked DNA important for lesion recognition in AlkC is stabilized by both the Ig-like domain and the N-terminal helical bundle, the conformation and topology of which is distinct from AlkD and other HLR proteins AlkD2 and AlkF/AlkG (Fig 5D and E) (Rubinson et al., 2008; Backe et al., 2013). In AlkD’s three-helical bundle (αA–αC), helix αB interacts with the damaged DNA strand from its position in the substrate (Mullins et al., 2015b). Attempts to trap ternary PfAlkC/1Ar-DNA/3mA and PfAlkC/THF-DNA/3mA complexes containing free 3mA nucleobase were unsuccessful, likely owing to the lack of base stacking at the sharp kink in the DNA, and suggesting that the nucleobase readily dissociates following N-glycosidic bond cleavage.
and is essential for binding affinity (Mullins et al., 2015a). In contrast, PfAlkC’s N-terminal helical bundle does not contact the damaged strand. PfAlkC’s helix αA and αB, which are broken into 2-helix segments to give αA’, αA, αB, and αB’, run antiparallel to one another and with opposite polarity with respect to BcAlkD so that αB does not contact the DNA. Instead, the N-terminal end of helix αA’ points toward the undamaged strand DNA, similar to the helix αC in both PfAlkC and BcAlkD (Fig 5D and E). As noted above, this region of AlkC may work together with the Ig-like domain to stabilize the bent DNA conformation.

The active sites of AlkC and AlkD are surprisingly different. AlkC contains a network of alternating charged residues along the damaged DNA backbone, whereas AlkD uses a Trp-Asp-Trp motif to cradle the backbone around the damaged nucleotide (Fig 5F and G). Each of the charged AlkC residues that span the damaged DNA backbone, whereas AlkD uses a Trp-Asp-Trp motif to contribute to catalysis, presumably by further stabilizing the development of positive charge on the deoxyribose as the N-glycosidic bond is broken (Mullins et al., 2015b; Parsons et al., 2016). In contrast, invariant Glu156 is positioned closer to the O4’ of the deoxyribose and is sandwiched between two arginine side chains from the ionic network, significantly lowering its predicted pKₐ (2.9) relative to that of Glu121 (4.3). We therefore speculate that Glu121 deprotonates the water nucleophile and that Glu156 plays more of a role in stabilizing positive charge that develops on the deoxyribose during base excision.

**AlkC catalyzes base excision of 3mC and 1mA from duplex DNA**

During our phylogenetic analysis, we noticed that most AlkC-containing bacteria lacked an AlkB ortholog, implying that these bacteria would be unable to repair 1-methyladenine (1mA) and 3-methylcytosine (3mC) lesions by oxidative demethylation. Out of 834 completely sequenced and annotated bacterial genomes that contain either an AlkB or AlkC ortholog, only 6% contained both proteins (Fig 6A). Given this distribution and the apparent specificity of AlkC for cationic methylbases, we tested the ability of purified BcAlkC (α) and PfAlkC (β) proteins to excise 1mA and 3mC from a double-stranded oligodeoxynucleotide substrate under single-turnover conditions. Compared to a no-enzyme control, BcAlkC and PfAlkC exhibited robust activity for 3mC and modest activity for 1mA, while AlkD had no activity toward either 3mC or 1mA (Fig 6B)

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**Figure 6. AlkC excises 3-methylcytosine (3mC) and 1-methyladenine (1mA) from DNA.**

A. Venn diagram of the numbers of bacterial species containing either AlkB (blue), AlkC (yellow), or both (green). B, C. Chemical structures and in vitro base excision of 3mC (B) and 1mA (C) from 25-mer double-stranded oligodeoxynucleotides. Representative denaturing electrophoresis gels show substrate (S) and product (P) after a 24-h incubation with either no enzyme (mock), BcAlkD, BcAlkC (AlkCα), or PfAlkC (AlkCβ). Plots show quantified time courses from three experiments (values are mean ± SD). D. 3mC may be repaired in bacteria by either AlkB-catalyzed oxidative demethylation or AlkC-catalyzed base excision. αKG, α-ketoglutarate.
and C, and Appendix Fig S4A and B). AlkC excision of 1mA was comparable to its weak activity for 7mG (Appendix Table S2), suggesting that 3mC is a more biologically relevant substrate for AlkC than is 1mA. Consistent with this, we were able to easily model 3mC, but not 1mA, into our crystal structure. Positioning 3mC in the active site places the N3-methyl group in van der Waals contact with Trp164 and the O2 oxygen in the Phe122/Glu121 pocket, whereas the N1-methyl group on 1mA sterically clashed with Trp164. 3mC and 1mA excisions were abrogated by Glu121Ala and Glu156Ala mutants and impaired by Trp164Ala substitution (Appendix Fig S4C and D), consistent with a catalytic role for the glutamates and a possible role for Trp164 in discrimination of different alkylated substrates. AlkC repair of these unusual lesions and the low percentage possibility that both AlkB and AlkC raise the interesting possibility that AlkC repairs 3mC and 1mA in bacteria that do not contain the AlkB oxidative demethylase (Fig 6D). Similarly, AlkB orthologs from the archaeon *Archaeoglobus fulgidus* and the archaea-related bacterium *Deinococcus radiodurans*, neither of which contain an AlkB homolog, are capable of 1mA and 3mC excision (Leiros et al, 2007; Moe et al, 2012). We did not observe 3mC or 1mA excision activity from ssDNA, nor did we observe excision of N1-methylguanine (1mG) or N3-methylthymine (3mT) (Appendix Fig S4E), all of which are substrates for AlkB (Falnes et al, 2002, 2004; Trewick et al, 2002; Delaney & Essigmann, 2004; Kolvisto et al, 2004), indicating that AlkB and AlkC are not strict functional orthologs.

**Conclusion**

Previous work defined the HLR domain as a new enzymatic scaffold for duplex DNA containing bulky lesions (Rubinson et al, 2008; Mullins et al, 2013). AlkC genes from *Bacillus anthracis* (NCBI WP_000521720.1), *Bacillus mycoides* (WP_003190106.1), *Bacillus weihenstephanensis* (WP_012261200.1), *Sphingobacterium* sp. 21 (WP_013668283.1), *P. fluorescens* SBW25 (WP_012723400.1), and *Streptomyces albus* (WP_003946460.1) were synthesized into expression vector pJ434 (DNA2.0). Constructs used in the genomic DNA base excision assay were overexpressed in *E. coli* C41(DE3) cells at 16°C overnight upon induction with 500 μM IPTG. For all other biochemical assays and X-ray crystallography, *P. fluorescens* alkC was subcloned into pBG100 (Vanderbilt Center for Structural Biology) encoding a cleavable N-terminal hexahistidine tag and overexpressed in *E. coli* HMS174(DE3) at 16°C overnight with 500 μM IPTG. Cells were lysed by sonication at 4°C in buffer A (50 mM Tris–HCl pH 8.5, 500 mM NaCl, and 10% (v/v) glycerol). AlkB orthologs were purified from soluble lysate by Ni-NTA (Qiagen) affinity chromatography by sequentially washing and eluting in buffer A containing 20 and 500 mM imidazole, respectively. Pooled fractions were digested with Rhinovirus 3C (PreCission) Protease overnight at 4°C to remove the N-terminal hexahistidine tag. Cleaved protein was diluted 10-fold in buffer B (50 mM Tris–HCl pH 8.5, 10% (v/v) glycerol, 2 mM DTT, and 0.1 mM EDTA) and purified over heparin sepharose (GE Healthcare) using a 50–1,000 mM NaCl linear gradient. The protein was further purified using gel filtration chromatography (Superdex 200, GE Healthcare) in buffer C (20 mM Tris–HCl pH 8.5, 150 mM NaCl, 10% (v/v) glycerol, 2 mM DTT, and 0.1 mM EDTA). Purified proteins were concentrated, flash-frozen, and stored at −80°C in buffer C.

PfAlkC mutants were constructed using the Q5 Site-Directed Mutagenesis Kit (New England BioLabs, Inc.). PfAlkC/C (residues 1–249) was generated by mutating the codon corresponding to residue 250 to a stop codon. Purification of PfAlkC/C mutants was conducted in the same manner as wild-type PfAlkC except that PfAlkC/C Glu121Ala was overexpressed in *E. coli* ArcticExpress(DE3) cells. Selenomethionine (SeMet)-incorporated PfAlkC/C was overexpressed and purified the same as wild-type PfAlkC, except overexpression was carried out in M9 minimal media supplemented with 0.4% (w/v) dextrose; 1 mM MgSO4; 0.1 mM CaCl2; 1 mg/l thiamine; 60 mg/l selenomethionine; 50 mg/l each of leucine,
isoleucine, and valine; and 100 mg/l each of phenylalanine, lysine, and threonine. Integrity of mutant proteins was verified by thermal denaturation, monitored by circular dichroism molar ellipticity at 222 nm on a Jasco J-810 spectropolarimeter using 7.5 μM protein in 50 mM HEPES pH 8.5, 100 mM KCl, and 10% (v/v) glycerol.

Base excision assays

Base excision using genomic DNA and oligonucleotide substrates was carried out as previously described (Mullins et al., 2013). Briefly, to measure release of methylbases from genomic DNA by AlkC orthologs, 5 μM protein was incubated with 10 μg of MNU-treated calf thymus DNA at 37°C for 1 h in 50 mM HEPES pH 7.5, 100 mM KCl, 5% (v/v) glycerol, 10 mM DTT, 2 mM EDTA and 0.1 mg/ml BSA in a 50 μl reaction and products quantified by HPLC-MS/MS. The activity of PfAlkC and mutants were tested in the same manner except that the assay was carried out at 21°C for 5 min in a 30 μl reaction consisting of 10 μg protein, 50 mM HEPES pH 8.5, 100 mM KCl, 5% (v/v) glycerol, and 6 μg of MNU-treated calf thymus DNA from a different batch. For oligonucleotide-based assays, 7mG, 1mA, 3mC, 1mG, or 3mT were incorporated into the presence of an 11-mer THF-DNA [d(TGTCCA(THF)GTCT)/d(GGCC)/d(GGCCCGCGCCCTTTGCCGCCGCCCG)] as described previously (Mullins et al., 2013) or by solid-phase synthesis by Midland Certified Reagent Company (1mA, 1mG, and 3mT) or ChemGenes Corporation (3mC) and annealed to the complementary strand in annealing buffer (10 mM MES pH 6.5 and 40 mM NaCl). The YTMA lesion was verified by ligand distances, coordination, and geometry using the CheckMyMetal web server (Zheng et al., 2014).

X-ray crystallography

Crystallization of the seven purified AlkC orthologs was first screened against a library of double-stranded oligonucleotides (Integrated DNA Technologies) ranging in length from 8 to 15 nucleotides and containing a centralized THF across from thymine. Diffracting crystals were obtained from the PfAlkC protein grown in the presence of an 11-mer THF-DNA [d(TGTCCA(THF)GTCT)/d(AAGACTTGGAC)] as described previously (Mullins et al., 2013), and the YTMA lesion was verified by ligand distances, coordination, and geometry using the CheckMyMetal web server (Zheng et al., 2014). Structural biology software was curated by SBGrid (Morin et al., 2013). Structure figures were prepared in PyMOL (Schrödinger, 2016). Conservation from 527 AlkC sequences was mapped onto the structure of PfAlkC using the ConSurf server (http://bental.tau.ac.il/new_ConSurfDB/). DNA geometric parameters were measured using Web 3DNA (Zheng et al., 2009). Structure-based pK_a calculations were carried out in Rosetta (Kilambi & Gray, 2012; Lyskov et al., 2013).

Data availability

Structure factors and coordinates were deposited in the Protein Data Bank under accession codes 5VHV (1aR complex) and 5V10 (THF complex), and the corresponding X-ray diffraction images deposited in the SBGrid Data Bank (Meyer et al., 2016).

Acknowledgements

This work was funded by the National Science Foundation (MCB-1517695 to B.F.E., CHE-1610721 to S.S.D.) and the National Institutes of Health (RO1 GM09625). Use of the Advanced Photon Source, an Office of Science User Facility operated for the U.S. Department of Energy Office of Science by Argonne National Laboratory, was supported by the U.S. Department of Energy (DE-AC02-06CH11357). Use of LS-CAT Sector 21 was supported by the Michigan Economic Development Corporation and the Michigan Technology Tri-Corridor (08SP1000817). The Advanced Light Source operated by Lawrence Berkeley National Laboratory on behalf of the Department of Energy, Office of Basic Energy Sciences, through the Integrated Diffraction Analysis Technologies (IDAT) program, was supported by DOE Office of Biological and Environmental Research. E.A.M. was supported by the Vanderbilt Training Program in Environmental Toxicology (T32ES07028). K.T.L. was supported by National Institutes of Health training program in chemical biology (T32 GM113770).
**Author contributions**

BFE conceived the project; RS, EAM, and BFE designed experiments; RS performed biochemical and structural experiments; EAM purified and analyzed activity of AlkC orthologs and generated preliminary PfAlkC-DNA crystals; KTL, performed biochemical and structural experiments; EAM purified and analyzed data; RS, EAM, and BFE wrote the manuscript.

**Conflicts of interest**

The authors declare that they have no conflict of interest.

**References**


Schroedinger L (2016) The PyMOL molecular graphics system, Version 1.7.4 Schrödinger, LLC.


Figure EV1. Comparison of DNA binding Ig folds.
The top two rows show orthogonal views of DNA-bound structures of Ig-like domains from three eukaryotic transcription factors Ndt80 (PDB 1MNN), NF-κB (PDB 1A3Q), and p53 (PDB 1TUP), compared to that of PfAlkC (this work). Topology diagrams of each structure are shown at the bottom and are colored the same as the crystal structures. The two β-sheets within the Ig core are colored gray (A, B, D, E, E', and H) and pale cyan (C, C', F, and G). Regions contacting the DNA are colored green (AB-loop), magenta (CC'-loop), orange (EF-loop), and red. Points of contact with the DNA in are marked with yellow stars in the topology diagram.
Figure EV2. Characterization of PfAlkC proteins used in this study.

A  SDS–PAGE of purified proteins, stained with Coomassie Blue.
B  Thermal denaturation of wild-type and PfAlkC mutants monitored by circular dichroism. The fraction of unfolded protein was expressed as the normalized molar ellipticity at 222 nm. Melting temperatures ($T_m$) were derived by fitting the data to the equation \( \text{fraction unfolded} = \frac{1}{1 + e^{(T_m - T_0)/k}} \), where $k$ denotes the cooperativity of the transition. Although PfAlkC$\Delta C$ has a lower $T_m$ than the full-length proteins, it is properly folded at 21°C at which biochemical assays were performed.
C  Electrophoretic mobility shift assay for PfAlkC mutants binding to 1 nM THF-DNA of the sequence $^{32}$P-d(GACCACTACACT(THF)ATTCCTAACAAC)/d(GTTGTTAGGAAT(T)AGTGTTAGTGGTC) in 25 mM HEPES (pH 7.5), 50 mM KCl, 5 mM DTT, 5% glycerol, and 0.05 mg/ml BSA at 21°C for 30 min. Concentrations of PfAlkC are shown at the top of the representative gels. Electrophoretic separation was carried out on a Novex™ TBE gel (ThermoFisher Scientific). Quantitation is plotted on the right, in which each value is the mean ± SD ($n = 3$). Equilibrium dissociation constants ($K_d$) were extracted by fitting the data to a two-state binding model.
Figure EV3. AlkC proteins contain an insertion that may stabilize DNA bend in the absence of Ig-like domain.

Two homology models for BcAlkC (center and right) are shown superimposed against the 1aR-DNA from the PfAlkC/1aR-DNA crystal structure (left). Only the N-terminal helical bundle (NTB, red), HEAT-like repeat 1 (HLR1, orange), and Ig-like domain (olive) are shown for clarity. A phenylalanine side chain at the N-terminus of helix αE in PfAlkC and in the 8-residue insertion of BcAlkC is shown in spheres and highlighted yellow in the sequence alignment at the bottom. The secondary structural elements from the three models and from a secondary structure prediction of BcAlkC are shown against the sequences for each protein. The homology models were generated in SWISS-MODEL (https://swissmodel.expasy.org/) using either the sequence alignment shown in Appendix Fig S1 (model 1) or generated by SWISS-MODEL (model 2). In both models, the insertion makes contacts to the 1aR strand, either as a loop (model 1) or as an N-terminal extension to helix αE (model 2).

Figure EV4. Crystallographic features of the active site.

A Na⁺ ion coordination in the 1aR-DNA structure. Na⁺ and water oxygens are shown as purple and red spheres, respectively. 1aR is magenta, and the opposite thymidine T6 is green. Superscripts in nucleotide labels refer to the 1aR-containing, damaged (D) strand or the opposite, undamaged (U) strand. Hydrogen bonds are shown as dashed lines with interatomic distances in Angstroms.

B Stereo-views of the PfAlkC/1aR-DNA active site. Active site residues are blue, and DNA is gold/magenta/green. Pentaerythritol propoxylate (PEP) sequestered from the solvent is in silver and superimposed against composite omit electron density contoured at 1σ. The putative catalytic water is shown as a red sphere. One arm of the PEP projects into the DNA kink between the 1aR and the flanking base pairs and thus may limit rotation of the 1aR ring back toward the DNA. The other two polymeric PEP arms project outward to solvent. The fourth PEP arm is not present; its terminal hydroxyl group forms a hydrogen bond to the displaced T6 thymine base. The position of the thymine is thus affected by the presence of the PEP in addition to its coordination by the Na⁺ ion. However, even in the absence of these stabilizing contacts, the T6 thymidine would be displaced into the minor groove as a result of the kink in the DNA. The space occupied by the PEP molecule would be occupied by the excised base both before and immediately after cleavage.
Figure EV5. Structure of the PfAlkC/THF-DNA complex and its comparison to PfAlkC/1aR-DNA.

A DNA glycosylase-catalyzed reaction together with chemical structures of intermediate and product mimetics used in this study.
B Annealed composite omit electron density contoured to 1σ is superposed onto the THF-DNA crystallographic model.
C Two views of the crystal structures, with protein in blue, DNA in gold, THF/1aR in magenta, and the thymidine opposite THF/1aR in green.
D DNA structures extracted from the complexes.
E Schematic of protein–DNA contacts. Contacts to the protein backbone are marked with an asterisk, and symmetry-related contacts are in parentheses. PfAlkC maintains similar contacts with the DNA in THF and 1aR complexes relative to the position of the DNA bend and not to the position of the 1aR/THF.
F Details of THF-DNA bound outside of the PfAlkC active site. Both complexes in the crystallographic asymmetric unit are shown. A PEG 4,000 molecule (white carbons) fills the void in the catalytic pocket in complex A.
G Superposition of the two protomers in the THF complex. Bold arrows highlight the difference in positions of the Ig-like domain relative to the HLR domain and in the two DNA conformations.
H Superposition of the two 1aR complexes.
I Superposition of one protomer from each of the THF (gold) and 1aR (cyan) complexes.
APPENDIX

Selective base excision repair of DNA damage by the non-base-flipping DNA glycosylase AlkC
Rongxin Shi, Elwood A. Mullins, Xing-Xing Shen, Kori T. Lay, Philip K. Yuen, Sheila S. David, Antonis Rokas, and Brandt F. Eichman*

Appendix Table S1. Crystallographic data collection and refinement statistics
Appendix Table S2. Base excision activities
Appendix Figure S1. AlkCα and AlkCβ sequence alignment
Appendix Figure S2. Substrate specificities of AlkC and AlkD
Appendix Figure S3. Glycosylase-induced DNA distortion
Appendix Figure S4. AlkC excision of 3mC and 1mA
Appendix References
### Appendix Table S1. Crystallographic data collection and refinement statistics

<table>
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$^a$ Statistics for the highest resolution shell are shown in parentheses.

$^b$ $R_{\text{free}}$ was determined from the 5% of reflections excluded from refinement.

$^c$ Riding hydrogen atoms were not included in no. atoms or avg. $B$-factors.

$^d$ In addition to water molecules, values for solvent include one PEG 4000 molecule in the THF structure and two Na+ ions, two pentaerythritol propoxylate, two glycerol, and three MES molecules in the 1aR structure.

$^e$ Equivalent isotropic $B$-factors were calculated in conjunction with TLS-derived anisotropic $B$-factors.
## Appendix Table S2. Base excision activities

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$^a$Values are single-turnover rate constants (s$^{-1}$), averaged from three independent measurements. Errors (SD) were between 10-20% for each.


$^c$n.d., none detected.
Appendix Figure S1. AlkCa and AlkCβ sequence alignment. Twenty selected sequences from four representative phyla (Actinobacteria, Bacteroidales, Firmicutes and Proteobacteria) were aligned using Clustal Omega and annotated using BoxShade (http://www.ch.embnet.org/software/BOX_form.html). Shaded residues have >50% sequence identity (black) and similarity (grey). Secondary structural elements identified from the PfAlkC crystal structure are shown below the sequences. Triangles designate PfAlkC residues important for base excision activity (red), nucleobase binding (orange) and for stabilizing the DNA backbone in the vicinity of the lesion (black).
Appendix Figure S2. Substrate specificities of AlkC and AlkD. A,B. Excision activity of BcAlkD and PfAlkC against oligodeoxynucleotides containing 7mG (A) and YTMA (B). Representative denaturing polyacrylamide gels show substrates (S) and hydroxide-nicked products (P) as a function of time. Quantification of the data is shown on the right; values are mean ± SD (n=3). The extra band below the 12-mer product in panel a corresponds to a nuclease contaminant in the AlkD preparation. The smearing in the AlkD-YTMA product band is a result of incompletely denatured GC-rich duplex DNA after hydroxide nicking.

C. Growth of B. anthracis wild-type (blue), ΔalkC (red), ΔalkD (green), and ΔalkCΔalkD (purple) in the presence of varying concentrations of yatakemycin (YTM). Bacillus anthracis ΔalkC cells were generated and their resistance to YTM assayed as described for ΔalkD (Mike et al, 2014; Mullins et al, 2015; Stauff & Skaar, 2009). Briefly, growth curves were obtained by growing cell cultures in the presence or absence of YTM and recording cell density every hour for 20 hours. Spot assays were performed by serial dilution of early-mid-log phase cells on LB plates prepared with or without YTM. Growth curves and spot assays were performed in triplicate.
Appendix Figure S3. Glycosylase-induced DNA distortion. DNA models from glycosylase-DNA co-crystal structures of PfAlkC/1aR-DNA (this work), BcAlkD/1aR-DNA (PDB 5KUB), and base-flipping human AAG/1,N6-ethenoadenine-DNA (PDB 1EWN) compared to unbound B-DNA (PDB 1BNA). The modified and partner nucleotides are colored magenta and green, respectively. Black arrows depict the helical axes.
Appendix Figure S4. AlkC excision of 3mC and 1mA. Representative denaturing electrophoresis gels show substrate (S) and NaOH-nicked abasic-DNA product (P) over a 24-hour incubation. Values in the plots are mean ± SD (n=3). A,B. 3mC-DNA (A) and 1mA-DNA (B) excision by no enzyme (mock), BcAlkD, BcAlkC, or PfAlkC. C,D. 3mC (C) and 1mA (D) excision by no enzyme (mock), wild-type PfAlkC (WT), or PfAlkC mutants (E121A, E156A, W164A, ΔC). E. 24-hour incubation of no enzyme [(−), mock], BcAlkC, or PfAlkC with double-stranded (ds) or single-stranded (ss) oligodeoxynucleotides containing a 3mC, 1mA, 1mG, or 3mT modification.
Appendix References


